

RELATIVE SUSCEPTIBILITY OF WOODY LANDSCAPE PLANTS TO JAPANESE BEETLE (COLEOPTERA: SCARABAEIDAE)

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Abstract. The Japanese beetle (*Popillia japonica* Newman) was introduced to a New Jersey nursery in 1916 and continues to spread across the United States and Canada. Adults attack foliage, flowers, and fruit of more than 300 species of plants; however, some plants are notably resistant. This paper summarizes data on plant susceptibility of woody plants to Japanese beetles collected from observations and controlled experiments. Resistance to Japanese beetle has been documented among species of maples (*Acer*) and birch (*Betula*) and among cultivars of crabapple (*Malus*), crapemyrtle (*Lagerstroemia*), and linden (*Tilia*). Production of certain plant odors, presence of secondary compounds in leaves, and leaf pubescence are factors affecting resistance to this insect. Host plant resistance is the most sustainable means of managing feeding damage or plant losses resulting from Japanese beetle adults. When suitable, incorporating Japanese beetle-resistant plants into new landscapes can reduce or eliminate the expense of replacing damaged plants or frequent insecticide applications.

Key Words. *Popillia japonica*; Japanese beetle; integrated pest management; host plant resistance.

The Japanese beetle (*Popillia japonica* Newman) is one of the most damaging pests of urban landscapes in the eastern United States. Yearly costs for management and mitigation of damage are estimated at US\$500 million (USDA/APHIS 2002). This scarab was introduced in 1916 to the eastern United States in infested nursery stock (Fleming 1972). At that time, entomologists were unaware of the pernicious nature of this species as evidenced from this quote, "While inspecting a nursery in southern New Jersey during August 1916, our attention was attracted by a scarabaeid feeding on the tips of *Crataegus*. ... Inasmuch as it was assumed to be a southern species, no particular attention to it was paid at that time" (Dickerson and Weiss 1918). In the presence of abundant grass and pasture land, and the apparent absence of natural enemies, the Japanese beetle flourished. Currently, this immigrant species partially infests or is established in all states east of the Mississippi River except Florida, and its range extends north into Canada (NAPIS 2003).

Adult Japanese beetles are broadly oval, 8 to 11 mm (about 0.5 in.) long, metallic green in color and have coppery-brown wing covers. Larvae are typical white grubs, C-shaped and cream colored, with three pairs of legs and a

light-brown head capsule (Fleming 1972). Japanese beetle has a 1-year life cycle, spending most of its life underground as a grub. A small grub, about 1.5 mm (0.06 in.) long, emerges from an egg laid 3 to 5 cm (1 to 2 in.) deep into moist soil, typically under turfgrass. Females alternate between periods of feeding and mating on host plants and oviposition. During her lifetime, a female will have 12 or more egg-laying bouts and produce 40 to 60 eggs. Once hatched, grubs feed on roots and will reach full size, about 32 mm (1.25 in.) long, by late summer (Fleming 1972). Management of grubs is accomplished with soil insecticides applied preventively (e.g., Merit or Mach2) before egg hatch, or curatively (e.g., Dylox or Sevin) after small grubs are present. Presence of grubs or grub damage may be associated with nearby plants infested with adults; however, females will disperse to find a suitable site for oviposition (Fleming 1972; Potter and Held 2002).

Japanese beetle adults are active from June through August in most of its geographic range. These beetles are day active and mate and feed concurrently on host plants. They can defoliate more than 300 species of woody and herbaceous plants in 79 plant families (Fleming 1972; Potter and Held 2002). Because of their mobility and gregarious habits, swarms of Japanese beetle continually infest and defoliate new plants during the growing season. These factors can complicate control of adults, especially when using short-residual insecticides such as pyrethroids. Certain systemic products delivered through soil or trunk injection are available for control of Japanese beetles on mature, established street and residential trees; however, that treatment may not be an option for newly installed landscape plantings.

Use of insecticides to manage Japanese beetle can be reduced if resistant plants are substituted for more susceptible ones in commercial and residential landscapes. Observations and controlled experiments indicate that certain plant species, and even cultivars of the same species, vary in susceptibility. For example, moderate or complete resistance to Japanese beetle feeding is documented for most evergreens, certain crabapples (*Malus*), lindens (*Tilia*), maples (*Acer*), birch (*Betula*), and crapemyrtles (*Lagerstroemia*) (Fleming 1972; Ranney and Walgenbach

1992; Spicer et al. 1995; Potter et al. 1998; Miller and Ware 1999; Pettis et al. 2004). There is no resistance to Japanese beetle among species or cultivars of rose (*Rosa*) (Potter et al. 1998; Held and Potter 2004).

Most of the information on host susceptibility to Japanese beetle originated from a landmark survey summarized by W.E. Fleming (1972). This publication has since gone out of print; however, the information remains relevant to urban horticulture because of the continued spread of Japanese beetles into the United States and Canada. In his review, Fleming (1972) established a damage rating system based on observations of plant damage noted for each plant species in his listing. This rating system is qualitative and assigns a relative rank to each species based on written and oral accounts of Japanese beetle feeding damage noted from 1920 through 1963, primarily in the New England area (Fleming 1972).

Additional laboratory and field evaluations of Japanese beetle susceptibility for certain horticulturally important taxa have been further investigated by contemporaries of Fleming. These subsequent studies compared the percentage of defoliation of field- or container-grown plants in a common garden type of experiment (e.g., Potter et al. 1998). Blocks of woody plants representing replicates of each cultivar or species were subject to defoliation by natural beetle populations during one or more years. Additional laboratory or controlled field experiments were also used to verify the results of field tests for crabapple (Ranney and Walgenbach 1992; Spicer et al. 1995), crapemyrtle (Pettis et al. 2004), and linden (Miller and Ware 1999). Besides the field observations on *Ulmus procera*, *U. rubra*, and *U. americana* in Fleming (1972), susceptibility of elm species is based on laboratory experiments with detached leaves or defoliation of plants caged with beetles (Miller et al. 2001). Discrepancies in seasonal results from multi-year field evaluations have been noted and are attributed to the relative abundance of adults from year to year (Fleming 1972; Potter et al. 1998).

The purpose of this paper is to provide landscape architects, professional landscape managers, and arborists a comprehensive list of woody plants and their relative susceptibility to Japanese beetle. Although the results of these resistance screenings were reported in scientific or extension publications, there has been no single source of host plant data for Japanese beetle since Fleming (1972). This paper has compiled the data from Fleming (1972) and amended it with data from recent experiments to produce a comprehensive record of plant susceptibility to Japanese beetle.

DESCRIPTION OF DATA PRESENTATION

Data are presented in table form, alphabetically by family, then scientific name. Tables 1 and 4 use a rating to indicate susceptibility. This rating is an adaptation of the system used by Fleming (1972). When a plant is designated "resistant," it

means observed plants were either never fed on or rarely fed on by Japanese beetles. "*" and "**" indicate plants on which feeding has been observed but is either occasional or light, respectively. "***" and "****" indicate plants that are commonly fed on by Japanese beetle, resulting in either moderate or extensive feeding damage, respectively. Plants with the latter two ratings will likely sustain considerable feeding damage or be completely defoliated if Japanese beetles are present.

Qualitative ratings for *Prunus serrulata*, *P. serotina*, and *P. virginiana* came from Fleming (1972), whereas all others were adapted from field defoliation data (Ranney and Walgenbach 1992). In the only field study with birch (*Betula*) species and cultivars, defoliation was 1% or less for all nine taxa, except for Himalayan birch (*B. jacquemontii*), which was 16% (Ranney and Walgenbach 1992). Based on these data, most birch are not preferred hosts (Table 1), except for Himalayan, European white, and gray, of which the latter two were ranked as more susceptible by Fleming (1972).

Tables 2 and 3 summarize resistance among cultivars of crapemyrtles and crabapples from field and laboratory experiments. Susceptibility ratings for crapemyrtle varieties are adapted from susceptibility rankings assigned by Pettis et al. (2004). The qualitative ratings assigned to crabapple cultivars (Table 3) were derived from three evaluations conducted in North Carolina (Ranney and Walgenbach 1992) and Kentucky (Spicer et al. 1995; Potter et al. 1998). Relative susceptibility of the 26 cultivars common to both sites was similar (Potter et al. 1998).

Crapemyrtle and crabapple species or cultivars are listed under headings indicating their relative susceptibility. As before, "resistant" indicates that observed plants were rarely fed on. For crabapples, only those with less than 10% defoliation in field studies were assigned to this rating. "Moderately resistant" means that beetle feeding was observed but light. Crabapples ranked as moderately resistant generally sustained 20% to 45% defoliation. Plants designated "moderately susceptible" will have noticeable damage by Japanese beetle corresponding to 50% to 70% defoliation for crabapple varieties. All plants considered "susceptible" will be extensively damaged or completely defoliated by Japanese beetle, equivalent to about 75% to 100% defoliation in the crabapple field studies (Ranney and Walgenbach 1992; Spicer et al. 1995; Potter et al. 1998).

Ratings for linden taxa (Table 4) were taken from observations in Fleming (1972), a 3-year field study of eight *Tilia* spp. in Kentucky (Potter et al. 1998), and Miller and Ware (1999), which combined laboratory feeding assays with leaves or leaf discs, with field defoliation data of 16 genotypes in Illinois. Ratings of linden were determined based on both studies; however, field defoliation data were used over laboratory results if there was any inconsistency between the relative rankings of the same variety.

Table 1. Relative susceptibility of deciduous and evergreen woody trees and shrubs to Japanese beetles.

Scientific name	Common name	Rating ^z	Scientific name	Common name	Rating ^z
Aceraceae			Cupressaceae		
<i>Acer negundo</i>	Boxelder	*	<i>Chamaecyparis lawsoniana</i>	Lawson white cedar	Resistant
<i>Acer palmatum</i>	Japanese maple	****	<i>Chamaecyparis obtuse</i>	Hinoki cypress	Resistant
<i>Acer platanoides</i>	Norway maple	****	<i>Chamaecyparis pisifera</i>	Sawara cypress	Resistant
<i>Acer pseudoplatanus</i>	Sycamore maple	**	<i>Chamaecyparis thyoides</i>	Atlantic white cedar	Resistant
<i>Acer rubrum</i>	Red maple	Resistant	<i>Juniperus chinensis</i>	Chinese juniper	*
<i>Acer saccharinum</i>	Silver maple	Resistant	<i>Juniperus communis</i>	Common juniper	*
<i>Acer saccharum</i>	Sugar maple	**	<i>Thuja occidentalis</i>	American arborvitae	*
			<i>Thuja orientalis</i>	Oriental arborvitae	*
Anacardiaceae			Ebenaceae		
<i>Cotinus coggygria</i>	Smoketree	*	<i>Diospyros virginiana</i>	Common persimmon	*
<i>Rhus copallina</i>	Flameleaf sumac	**			
<i>Rhus typhina</i>	Staghorn sumac	*	Ericaceae		
Aquifoliaceae			<i>Kalmia latifolia</i>	Mountain laurel	Resistant
<i>Ilex aquifolium</i>	English holly	Resistant	<i>Rhododendron catawbiense</i>	Catawba rhododendron	*
<i>Ilex cornuta</i>	Chinese holly	Resistant	<i>Rhododendron maximum</i>	Rosebay rhododendron	*
<i>Ilex crenata</i>	Japanese holly	Resistant	<i>Rhododendron periclymenoides</i>	Pinxterbloom azalea	Resistant
<i>Ilex opaca</i>	American holly	Resistant	<i>Rhododendron viscosum</i>	Swamp azalea	*
<i>Ilex verticillata</i>	Winterberry holly	*	Fabaceae		
Berberidaceae			<i>Albizia julibrissin</i>	Mimosa	Resistant
<i>Berberis thunbergii</i>	Japanese barberry	**	<i>Cercis canadensis</i>	Eastern redbud	Resistant
Betulaceae			<i>Cercis chinensis</i>	Chinese redbud	Resistant
<i>Alnus glutinosa</i>	Black alder	***	<i>Robinia pseudoacacia</i>	Black locust	*
<i>Betula ermanii</i>	Erman's birch	Resistant	<i>Sophora japonica</i>	Japanese pagoda tree	*
<i>Betula jacquemontii</i>	Himalayan birch	***	<i>Wisteria sinensis</i>	Chinese wisteria	***
<i>Betula nigra</i>	River birch	*	Fagaceae		
<i>Betula nigra</i> 'Heritage'	Heritage river birch	*	<i>Castanea crenata</i>	Japanese chestnut	**
<i>Betula papyrifera</i>	Paperbark birch	Resistant	<i>Castanea dentata</i>	American chestnut	****
<i>Betula pendula</i>	European white birch	***	<i>Fagus grandifolia</i>	American beech	**
<i>Betula platyphylla</i> var. <i>japonica</i> 'Whitespire'	Asian Whitespire birch	Resistant	<i>Fagus sylvatica</i>	European beech	**
<i>Betula platyphylla</i> var. <i>szechuanica</i>	Asian Szechuan birch	Resistant	<i>Quercus alba</i>	White oak	*
<i>Betula populifolia</i>	Gray birch	****	<i>Quercus coccinea</i>	Scarlet oak	*
<i>Corylus americana</i>	American filbert	*	<i>Quercus falcata</i>	Southern red oak	*
<i>Corylus colurna</i>	Turkish filbert	**	<i>Quercus prinus</i>	Chestnut oak	**
Bignoniaceae			<i>Quercus palustris</i>	Pin oak	***
<i>Catalpa bignonioides</i>	Southern catalpa	***	<i>Quercus rubra</i>	Red oak	*
Buxaceae			<i>Quercus stellata</i>	Post oak	*
<i>Buxus sempervirens</i>	Common boxwood	Resistant	<i>Quercus velutina</i>	Black oak	*
Calycanthaceae			Ginkgoaceae		
<i>Calycanthus floridus</i>	Carolina allspice	Resistant	<i>Ginkgo biloba</i>	Maidenhair tree	*
Caprifoliaceae			Hamamelidaceae		
<i>Lonicera fragrantissima</i>	Winter honeysuckle	Resistant	<i>Hamamelis virginiana</i>	Witch hazel	*
<i>Lonicera japonica</i>	Japanese honeysuckle	*	<i>Liquidambar styraciflua</i>	American sweetgum	*
<i>Sambucus canadensis</i>	American elder	*	Hippocastaneae		
<i>Symphoricarpos albus</i>	Snowberry	Resistant	<i>Aesculus hippocastanum</i>	Horsechestnut	****
<i>Symphoricarpos orbiculatus</i>	Buckbrush	Resistant	<i>Aesculus parviflora</i>	Bottlebrush buckeye	***y
<i>Viburnum dentatum</i>	Arrowwood	***	Hypericaceae		
<i>Viburnum opulus</i>	European cranberry bush	*	<i>Hypericum perforatum</i>	Common St. Johnswort	***y
<i>Weigela florida</i>	Weigela	**	Juglandaceae		
Celastraceae			<i>Carya glabra</i>	Pignut hickory	*
<i>Celastrus scandens</i>	American bittersweet	Resistant	<i>Carya ovata</i>	Shagbark hickory	*
<i>Euonymus alatus</i>	Burning bush	*	<i>Carya tomentosa</i>	Mockernut hickory	**
<i>Euonymus fortunei</i>	Wintercreeper euonymus	Resistant	<i>Juglans cinerea</i>	Butternut	*
Clethraceae			<i>Juglans nigra</i>	Black walnut	****
<i>Clethra alnifolia</i>	Summersweet clethra	***y	Lauraceae		
Cornaceae			<i>Lindera benzoin</i>	Common spicebush	***
<i>Cornus florida</i>	Flowering dogwood	Resistant	<i>Sassafras albidum</i>	Common sassafras	****
			Loganiaceae		
			<i>Buddleia davidii</i>	Butterfly-bush	***y
			<i>Buddleia alternifolia</i>	Alternate-leaf butterfly-bush	***y

Table 1. Relative susceptibility of deciduous and evergreen woody trees and shrubs to Japanese beetles.

Scientific name	Common name	Rating ^z	Scientific name	Common name	Rating ^z
Lythraceae			Rubiaceae		
<i>Lagerstroemia</i> (see Table 2)	Crapemyrtle	****y	<i>Rosa</i> spp. and hybrids	Roses	****y
Magnoliaceae			<i>Sorbus americana</i>	American mountain ash	****
<i>Liriodendron tulipifera</i>	Tulip poplar	Resistant	<i>Spiraea trilobata</i>	Three-lobed spirea	**
<i>Magnolia grandiflora</i>	Southern magnolia	*y	<i>Spiraea</i> × <i>vanhoutei</i>	Vanhoutte spirea	**
<i>Magnolia</i> × <i>soulangiana</i>	Saucer magnolia	Resistant	Rutaceae		
<i>Magnolia virginiana</i>	Sweetbay magnolia	Resistant	<i>Cephalanthus occidentalis</i>	Buttonbush	****y
Moraceae			<i>Gardenia jasminoides</i>	Gardenia	*
<i>Ficus carica</i>	Common fig	Resistant	Salicaceae		
<i>Ficus elastica</i>	Indian rubber tree	*	<i>Populus alba</i>	White poplar	Resistant
<i>Morus rubra</i>	Red mulberry	Resistant	<i>Populus alba pyramidalis</i>	Bolleana poplar	Resistant
Musaceae			<i>Populus nigra italica</i>	Lombardy poplar	****
<i>Musa</i> × <i>paradisical</i>	French plantain	*	<i>Salix babylonica</i>	Babylon weeping willow	**
Myricaceae			<i>Salix discolor</i>	Pussy willow	****
<i>Myrica pensylvanica</i>	Northern bayberry	**	Saxifragaceae		
Nyssaceae			<i>Deutzia gracilis</i>	Deutzia	**y
<i>Nyssa sylvatica</i>	Tupelo	**	<i>Hydrangea arborescens</i>	Smooth hydrangea	Resistant
Oleaceae			<i>Hydrangea paniculata</i>	Panicle hydrangea	Resistant
<i>Forsythia</i> × <i>intermedia</i>	Border forsythia	Resistant	<i>Hydrangea petiolaris</i>	Climbing hydrangea	*
<i>Forsythia suspensa</i> var. <i>sieboldii</i>	Weeping forsythia	Resistant	<i>Philadelphus coronaries</i>	Mockorange	Resistant
<i>Fraxinus americana</i>	White ash	Resistant	Simaroubaceae		
<i>Fraxinus pennsylvanica</i>	Green ash	Resistant	<i>Ailanthus altissima</i>	Tree of Heaven	*
<i>Ligustrum ovalifolium</i>	California privet	**	Staphyleaceae		
<i>Ligustrum vulgare</i>	Common privet	*	<i>Staphylea trifolia</i>	American bladdernut	Resistant
<i>Syringa</i> × <i>persica</i>	Persian lilac	Resistant	Styracaceae		
<i>Syringa vulgaris</i>	Common lilac	Resistant	<i>Halesia tetraptera</i>	Carolina silverbell	***
Pinaceae			Taxaceae		
<i>Abies concolor</i>	Balsam fir	Resistant	<i>Taxus baccata</i>	English yew	Resistant
<i>Larix deciduas</i>	European larch	**	<i>Taxus brevifolia</i>	Western yew	Resistant
<i>Picea abies</i>	Norway spruce	Resistant	<i>Taxus canadensis</i>	Canada yew	Resistant
<i>Picea orientalis</i>	Oriental spruce	Resistant	<i>Taxus cuspidate</i>	Japanese yew	Resistant
<i>Pinus sylvestris</i>	Scotch pine	Resistant	Taxodiaceae		
<i>Pinus virginiana</i>	Virginia pine	*	<i>Cryptomeria japonica</i>	Cryptomeria	*
<i>Pseudotsuga menziesii</i>	Douglasfir	Resistant	<i>Taxodium distichum</i>	Baldcypress	***
<i>Tsuga canadensis</i>	Hemlock	Resistant	Tiliaceae (see Table 4)		
Platanaceae			Ulmaceae		
<i>Platanus</i> × <i>acerifolia</i>	London planetree	****	<i>Ulmus americana</i>	American elm	****
<i>Platanus occidentalis</i>	American planetree	***	<i>Ulmus changii</i>		****
Rosaceae			<i>Ulmus lanceaefolia</i>		**
<i>Chaenomeles japonica</i>	Japanese flowering quince	**	<i>Ulmus procera</i>	English elm	****
<i>Crateagus laevigata</i>	English hawthorn	**	<i>Ulmus prunifolia</i>		**
<i>Crateagus monogyna</i>	Singleseed hawthorn	**	<i>Ulmus pseudopropinqua</i>		**
<i>Exochorda racemosa</i>	Common pearlbrush	**	<i>Ulmus rubra</i>	Slippery elm	**
<i>Malus</i> (see Table 3)			<i>Ulmus taihangshanensis</i>		****
<i>Prunus</i> × <i>cistena</i>	Purpleleaf sandcherry	****	<i>Ulmus wallichiana</i>		****
<i>Prunus sargentii</i>	Sargent cherry	****	Verbenaceae		
<i>Prunus serotina</i>	Black cherry	****	<i>Callicarpa dichotoma</i>	Purple beautyberry	Resistant
<i>Prunus serrulata</i>	Oriental cherry	**	<i>Lantana camara</i>	Lantana	Resistant
<i>Prunus serrulata</i> 'Kwanzan'	Kwanzan oriental cherry	**	*Plants designated "resistant" are never fed on or rarely fed on by Japanese beetles. "*" and "**" indicate plants on which feeding has been observed but is either occasional or light, respectively. "****" and "*****" indicate plants that are commonly fed on by Japanese beetle, resulting in either moderate or extensive feeding damage, respectively.		
<i>Prunus serrulata</i> 'Mt. Fuji'	Mt. Fuji oriental cherry	****	†Flowers of these species are also fed on by Japanese beetles.		
<i>Prunus serrulata</i> 'Tai Haku'	Tai Haku oriental cherry	**			
<i>Prunus subhirtella</i>	'Autumnalis Rosea'	****			
<i>Prunus virginiana</i>	Common chokecherry	**			
<i>Prunus</i> × <i>incamp</i> 'Okame'	Okame cherry	****			
<i>Prunus</i> × <i>yedoensis</i> 'Afterglow'	Afterglow Yoshino cherry	**			
<i>Prunus</i> × <i>yedoensis</i> 'Akebono'	Akebono Yoshino cherry	**			
<i>Pyracantha coccinea</i>	Firethorn	*			
<i>Pseudocdonia sinensis</i>	Chinese quince	**			
<i>Pyrus communis</i>	Pear	*			

Table 2. Relative susceptibility of crapemyrtles to Japanese beetles.

Resistant		
'Acoma'	'Pocomoke'	
Moderately resistant		
'Biloxi'	'Cordon Bleu'	'Potomac'
'Catawba'	'Lipan'	'Sioux'
'Chicksaw'	'Muskogee'	'Tuskegee'
'Choctaw'	'Osage'	'Wichita'
'Comanche'	'Pink Velour'	
Moderately susceptible		
'Apalachee'	'Hope'	'Seminole'
'Byers Standard Red'	'Hopi'	'Tonto'
'Byers Wonderful White'	'Miami'	'Tuscarora'
'Carolina Beauty'	'Natchez'	'Velma's Royal Delight'
'Centennial'	'Ozark Springs'	'Victor'
'Centennial Spirit'	'Pecos'	'William Toovey'
'Dynamite'	'Powhatan'	'World's Fair'
'Hardy Lavender'	'Raspberry Sundae'	'Yuma'
Susceptible		
'Red Rocket'	'Regal Red'	

Table 3. Relative susceptibility of crabapples to Japanese beetles.

Resistant		
<i>Malus baccata</i> Jackii	'Harvest Gold'	'Strawberry Parfait'
<i>Malus hupehensis</i>	'Jewelberry'	
'Golden Raindrops'	'Louisa'	
Moderately resistant		
'Adirondack'	<i>Malus halliana</i> var. <i>parkmanii</i>	'Red Jewel'
'Baskatong'	<i>Malus tschonoski</i>	<i>Malus sargentii</i>
'Bob White'	'Madonna'	'Sentinel'
'Brandywine'	'Molten Lava'	'Silver Moon'
'Callaway'	'Naragansett'	'Snowdrift'
'Centurion'	'Ormiston Roy'	'Sugar Tyme'
'Christmas Holly'	'Professor Sprenger'	'Wintergold'
'David'	'Profusion'	<i>Malus × zumi</i> 'Calocarpa'
'Dobloons'	'Ralph Shay'	<i>Malus × zumi</i> 'Winter'
'Gem'		
'Edna Mullins'	'Red Jade'	
Moderately susceptible		
'Adams'	'Indian Magic'	'Ruby Luster'
'Beverly'	'Indian Summer'	'Selkirk'
'Candymint Sargent'	'Mary Potter'	'Sinai Fire'
'Coralburst'	'Pink Princess'	'Snow Magic'
'Donald Wyman'	'Purple Prince'	'Tina'
<i>Malus floribunda</i>	'Red Baron'	'White Angel'
'Henningii'	'Robinson'	
Susceptible		
<i>Malus baccata</i>	'Liset'	'Royalty'
'Dolgo'	'Radiant'	'Velvet Pillar'
'Hopa'	'Red Splendor'	'Weeping Candied Apple'

Table 4. Relative susceptibility of lindens to Japanese beetles evaluated in laboratory or field experiments.

Scientific name	Common name/cultivar	Rating ^z
<i>Tilia amurensis</i>		**
<i>Tilia americana</i>		*
<i>Tilia americana</i>	'Legend'	**
	'Redmond'	***
<i>Tilia caroliniana</i>		*
<i>Tilia chinensis</i>		**
<i>Tilia cordata</i>	'Chancellor'	***
	'Fairview'	**
	'Glenleven'	**
	'Greenspire'	****
	'Olympic'	****
	'Prestige'	***
<i>Tilia</i> × <i>euchlora</i>		***
<i>Tilia heterophylla</i>	'Continental Appeal'	**
<i>Tilia japonica</i>		*
<i>Tilia maximowicziana</i>		**
<i>Tilia mongolica</i>		**
<i>Tilia oliveri</i>		*
<i>Tilia orbicularis</i>		***
<i>Tilia petiolaris</i>	Pendent silver linden	*
<i>Tilia platyphyllos</i>	Largeleaf linden	***
<i>Tilia platyphyllos</i>	'Parade'	*
<i>Tilia tomentosa</i>		**
<i>Tilia tomentosa</i>	'Erecta'	**
<i>Tilia tomentosa</i>	'Sterling'	**
<i>Tilia</i> sp.	'Sundance'	*

^z "*" and "***" indicate plants on which feeding has been observed but is either occasional or light, respectively. "****" and "*****" indicate plants that are commonly fed on by Japanese beetle, resulting in either moderate or extensive feeding damage, respectively.

DISCUSSION

Susceptibility of plants to Japanese beetle should be one factor, among many, considered when selecting plants, particularly long-lived woody plants, for residential and commercial landscapes. Resistance of one plant species to Japanese beetle does not necessarily imply resistance to other plant-feeding insects or plant pathogens (Smitley and Peterson 1993; Pettis et al. 2004). For example, the crapemyrtle varieties 'Tonto' and 'Tuscarora' are moderately susceptible to Japanese beetle, but the same varieties are resistant to metallic flea beetles (*Altica* spp.), an important pest of crapemyrtle in production (Pettis et al. 2004).

Resistance of woody host plants to Japanese beetle is probably mediated by the presence or absence of deterrent compounds found in the foliage (Keathley et al. 1999; Potter and Held 2002). Control products containing certain plant extracts, such as neem (azadirachtin), can effectively deter feeding in laboratory choice tests (Ladd et al. 1978; Held et al. 2001) but often fail to provide similar protection when tested on whole plants in the field (Harper and Potter 1994; Witt et al. 1999). Abundant field populations, however, will

reduce the efficacy of both conventional and botanical insecticides because of additional adults re-infesting treated plants.

Elms and lindens are considered preferred hosts for the Japanese beetle. Among elms, only *U. lancefolia* and *U. prunifolia* were slightly less susceptible than the other species (Miller et al. 2001). Although no lindens are resistant, varieties such as 'Parade', 'Legend', and 'Sterling' appear to be less susceptible (Potter et al. 1998). Moderate to dense leaf pubescence may be an important factor in susceptibility of linden and elm to Japanese beetle. For example, foliage of *T. platyphyllos* 'Parade', *T. tomentosa* 'Sterling', and *U. lamellosa* have heavy pubescence and is less preferred by Japanese beetle (Potter et al. 1998; Miller and Ware 1999; Miller et al. 2001). Conversely, certain plants, such as species of *Ilex* and *Rhododendron*, with waxy or glossy foliage are also resistant to Japanese beetle (Fleming 1972; Keathley et al. 1999).

Plants with purplish or deep red foliage (e.g., 'Crimson King' Norway maple) are often observed to sustain more damage by Japanese beetle than green-leaved cultivars (Rowe et al. 2002). Foliage color alone, however, does not account for these differences. When two artificial ficus trees with foliage painted either green or purple are placed side by side in the field, significantly more beetles land on the green-leaved plants (Rowe et al. 2002). Flower color, however, does influence susceptibility of flowering plants to Japanese beetle. Rose varieties with yellow or white flowers are more likely to be attacked than those with darker-colored blooms (Held and Potter 2004).

Resistance of certain plants to Japanese beetle also depends on the production of attractive volatile compounds following damage by Japanese beetle or other plant-feeding insects (Loughrin et al. 1995). Japanese beetles use a wide range of floral and fruitlike compounds to locate a host plant (Fleming 1972; Loughrin et al. 1995, 1996). Laboratory tests show that Japanese beetle often cannot discriminate among foliage of plants that differ in susceptibility in the field (Loughrin et al. 1995, 1996). This finding indicates that Japanese beetles are attracted to plants regardless of their status as a host (Potter and Held 2002). However, if susceptible plants suffer feeding damage, they produce an array of attractive volatiles that serve as aggregation stimulants for Japanese beetle (Loughrin et al. 1995, 1996). Therefore, a susceptible plant in the field may not be inherently more attractive, but, if damaged, these plants produce the volatiles that recruit Japanese beetles much like sharks attracted to a blood trail in the water.

Host plant resistance is the most sustainable means of managing feeding damage or plant losses resulting from Japanese beetle adults. Landscape designers in states on the front of this insect's range expansion should consider incorporating resistant plants into residential, commercial, and municipal landscapes as well as any other long-term

plantings. This approach can reduce the economic and environmental costs associated with the repeated use of insecticides to prevent or reduce damage to urban landscapes in the future.

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Zusammenfassung. Der japanische Käfer *Popillia japonica* Newman wurde 1916 in eine Baumschule in New Jersey eingeführt und breitet sich nach Norden und Osten quer durch die Vereinigten Staaten aus. Die Erwachsenen attackieren Blätter, Blüten und Früchte von mehr als 300 Arten, dennoch sind einige resistent geblieben. Dieses Papier erfasst die Daten der für diesen Käfer anfälligen Gehölze, die aus Beobachtungen und kontrollierten Experimenten stammen. Eine Resistenz gegenüber Jap. Käfer wurde bei einigen Ahornarten, Birken, einigen Kultivaren von *Malus*, *Lagerstroemia* und *Tilia* dokumentiert. Die Produktion bestimmter Pflanzengerüche, Präsenz sekundärer Inhaltstoffe in den Blättern und behaarte Blätter sind Faktoren, die die Resistenz gegenüber diesem Insekt beeinflussen. Die Resistenz der Wirtspflanze ist die meist-zuerhaltene Mittel um Fraßschäden oder Pflanzenverluste durch den Käfer zu managen. Wenn möglich kann die Inkorporation resistenter Pflanzen in neue Landschaften die Ausgaben für die Ersatzpflanzungen beschädigter Gehölze reduzieren und die Insektizidanwendungen minimieren.

Resumen. El escarabajo japonés, *Popillia japonica* Newman, fue introducido a New Jersey en 1916, y continúa esparciéndose al norte y este a través de los Estados Unidos. Los adultos atacan el follaje, flores y frutos de más de 300 especies de plantas. Sin embargo, algunas plantas son notablemente resistentes. Este reporte resume los datos sobre la susceptibilidad de plantas leñosas a los escarabajos japoneses colectados de observaciones y experimentos controlados. La resistencia al escarabajo japonés ha sido documentada entre especies de maples (*Acer*), abedules (*Betula*) y entre cultivares de manzanos (*Malus*), astronómicas (*Lagerstroemia*) y tilos (*Tilia*). La producción de ciertos olores

por las plantas, presencia de componentes secundarios en las hojas y pubescencia de las mismas son factores que afectan la resistencia a estos insectos. La resistencia de las plantas hospederas es el medio más sustentable de manejo del daño o pérdida de plantas resultante de los escarabajos japoneses adultos. Cuando se desea, la incorporación de plantas resistentes en nuevos escenarios puede reducir o eliminar los gastos de reemplazo de plantas dañadas o las aplicaciones frecuentes de insecticidas.

Résumé. Le scolyte japonais (*Popillia japonica* Newman) a été introduit dans une pépinière du New Jersey en 1916 et continue depuis de s'étendre vers le nord et l'est des États-Unis. Les adultes attaquent le feuillage, les fleurs et les fruits de plus de 300 espèces de plantes, mais certaines plantes demeurent résistantes de manière notable. Cet article fait un résumé des données recueillies sur la susceptibilité des plantes ligneuses aux scolytes japonais, et ce à partir de d'observations et d'expériences contrôlées. La résistance au scolyte japonais a été documentée parmi des espèces des genres *Acer*, *Betula* et sur certains cultivars de *Malus*, de *Lagerstroemia* et de *Tilia*. La production de certains odeurs par les plantes, la présence de composés secondaires dans les feuilles et la pubescence foliaire sont des facteurs qui affectent la résistance à cet insecte. La résistance de la plante hôte est le moyen le plus efficace pour gérer les dommages causés par l'alimentation ou encore les pertes de végétaux résultants des scolytes japonais adultes. Lorsque cela est possible, l'incorporation de plants résistants au scolyte japonais dans les nouveaux aménagements peut réduire ou éliminer les dépenses pour le remplacement de végétaux endommagés ou les applications fréquentes d'insecticide.